How do variability&size control affect population growth?



• Assume here a constant environment, and will not consider "bet-hedging" scenarios *e.g., Balaban et al., Science (2004)*

How do variability&size control affect population growth?



Jie Lin

Ethan Levien

Single-cell variability: *Gaining* from noise?



independent generation time model

$$2\int_{0}^{\infty}e^{-\Lambda_{p} au}f(au)d au=1$$
 Powell,
Microbiology, 1956
Key Assumption:

no correlation in mother-daughter generation time

Result: variability enhances the population growth

Noise-driven growth rate gain in clonal **PNAS**, 2016 cellular populations

Mikihiro Hashimoto^a, Takashi Nozoe^a, Hidenori Nakaoka^a, Reiko Okura^a, Sayo Akiyoshi^a, Kunihiko Kaneko^{a,b}, Edo Kussell^{c,d}, and Yuichi Wakamoto^{a,b,1}

Noise and Epigenetic Inheritance of Single-Cell *Current Biology, 2016* **Division Times Influence Population Fitness**

Bram Cerulus, Aaron M. New, Ksenia Pougach, Kevin J. Verstrepen

Simplified scenario: no growth rate variability

Cell size is regulated — the population growth rate = the volume growth rate

$$\frac{dV}{dt} = \sum_{i=1}^{N(t)} \frac{dv_i}{dt} = \sum_{i=1}^{N(t)} \lambda_0 v_i = \lambda_0 V \qquad \qquad \Lambda_p = \Lambda_v = \lambda_0$$



Statistics on lineage trees



$$2 \int_{0}^{\infty} e^{-\Lambda_{p}\tau} f_{0}(\tau) d\tau = 1 \qquad \qquad f_{2}(\tau) = 2(1 - e^{-\Lambda_{p}\tau}) f_{0}(\tau)$$

Lin and Amir, *Cell Systems* (2017); Levien, Kondev and Amir, Biorxiv:680066 (2019) See also: Lebowitz and Rubinow, *Journal of Mathematical Biology* (1974)





The book: What If? Serious Scientific Answers to Absurd Hypothetical Questions



e than of living ones?
e than of living ones?
Emily Dunham
n
X
(

Growth rate and doubling time variability

- Two sources of noise
- σ_T = affects **generation time** noise
- σ_{λ} = growth rate variability
- Growth rate fluctuations are often small: CV of 6-8 %
- Generation time variability : CV of 20-30%

$$\Lambda_p(\sigma_t, \sigma_\lambda, \alpha) \approx \Lambda_p(\#, \sigma_\lambda, \#)$$

(as long as size is controlled!)





Only stochasticity in growth rate matters



Can we test this?



Lin and Amir, Cell Systems (2017)

Generalizations: growth rate correlations



Conditional probability distribution

$$h(\tau'|\tau)$$

$$N(t) = A(\tau)e^{\Lambda_p t} \Longrightarrow A(\tau)e^{\Lambda_p t} = 2\int_0^\infty A(\tau')e^{\Lambda_p(t-\tau)}h(\tau'|\tau)d\tau'$$

Lin and Amir, arXiv: 1806.02818

Single-cell variability: *Gaining/Losing* from noise



$$\Lambda_p = 1 - \left(1 - \frac{\ln(2)}{2} \frac{1+a}{1-a}\right) \sigma_\lambda^2$$
$$a_c = \frac{2 - \ln(2)}{2 + \ln(2)} \approx 0.5$$

Lin and Amir, arXiv: 1806.02818

Single-cell variability: origins and fitness advantage





e.g. bacterial persistence Gene expression patterns Plasmids

Generation time and cell size



Deterministic

e.g. aging in yeast Partitioning of efflux pumps in bacteria Growth of *M. tuberculosis*

Lin, Min and Amir, *Physical Review Letters* (2019)

Martins and Locke, Current Opinion in Microbiology (2015)

Another mechanism of phenotypic heterogeneity: Asymmetric protein segregation

Deleterious proteins

Cell Systems



Coelho et al., Current Biology 2013

Beneficial proteins



Bergmiller et al., Science 2017



Asymmetric Damage Segregation Constitutes an Emergent Population-Level Stress Response

Søren Vedel, ^{1,2,*} Harry Nunns, ^{1,3} Andrej Košmrlj,⁴ Szabolcs Semsey,¹ and Ala Trusina^{1,*}

Under what conditions does asymmetric protein segregation enhance the population fitness?

Mathematical model



(iii) Asymmetric segregation of protein

(ii) Protein concentration determines the single-cell growth rate







Jiseon Min

Jie Lin

Lin, Min and Amir, PRL (2019)

Solution for small asymmetry:



Solution for total asymmetry:

Leading term changes sign at the inflection point S_c

 $\lambda(\sigma)$

damage



Graphical interpolation



Result of graphical interpolation: Phase transitions!

- Control parameter: the protein accumulation rate, S
- Order parameter: the optimal asymmetry degree, a_c



Acknowledgments



Felix Barber (yeast size control) <u>Yipei Guo (bacterial evolution)</u> Felix Wong (bacterial mechanics) <u>Po-Yi Ho (cell cycle regulation)</u> <u>Dr. Ethan Levien (bet hedging)</u> <u>Dr. Jie Lin (population growth)</u> <u>Jiseon Min (damage partitioning)</u> Paul Dieterle (collective dynamics)

David Nelson, Ethan Garner, Andrew Murray, Nancy Kleckner (Harvard) Bree Aldridge and Michelle Logsdon (Tufts) Lars Renner (Dresden) Sven van Teefelen (Pasteur) Ilya Soifer (Calico) Naama Barkai (Weizmann) Nathalie Balaban (Hebrew University)